CONTENTS

AGRICULTURAL ENGINEERING

97-102 Distribution of stress in greenhouses frames estimated by aerodynamic coefficients of Brazilian and European standards
José Gabriel Vieira Neto, Julio Soriano

ANIMAL SCIENCE AND PASTURES

103-108 Conjugated linoleic acid of dairy foods is affected by cows’ feeding system and processing of milk
Juan Pablo Aviles Ruiz, Marcelo Wladimir Alonzo, Manuel Delgado Pertíñez

CROP SCIENCE

109-114 Initial horticultural performance of nine ‘Persian’ lime selections grafted onto Swingle citrumelo
Magno Guimarães Santos, Walter dos Santos Soares Filho, Eduardo Augusto Girardi, Abielmon da Silva Gesteira, Orlando Sampaio Passos, Claudia Fortes Ferreira

ENTOMOLOGY

115-124 Resistance mechanisms of sugarcane cultivars to spittlebug Mahanarva fimbriolata
Leila Luci Dinardo-Miranda, Juliano Vilela Fracasso, Dilermando Perecin, Mariana Cecília de Oliveira, Diego Olimpio Peçoto Lopes, Tiago Sabongi Izeppi, Ivan Antonio dos Anjos

FOOD SCIENCE AND TECHNOLOGY

125-133 Evaluation of natural preservatives in combination with acid whey for use in fermented sausage
Karolina Maria Wójciak, Zbigniew Józef Delatowski

GENETICS AND PLANT BREEDING

134-141 Genetic diversity reduction in improved durum wheat cultivars of Morocco as revealed by microsatellite markers
Fatima Henkrar, Jamal El-Haddouar, Hassan Ouababou, Nasserelhaq Nsarellah, Driss Iraqi, Najib Bendaaou, Sripada Mahabala Usupa

142-149 Genome selection in fruit breeding: application to table grapes
Alexandre Pio Viana, Marcos Deon Viilela de Resende, Summaira Riaz, Michael Andrew Walker

PLANT PATHOLOGY

150-158 Differential leaf gas exchange performance of mango cultivars infected by different isolates of Ceratocystis fimbriata
Wilka Messner da Silva Bispo, Leonardo Araújo, Wiler Ribas Moreira, Leandro de Castro Silva, Fabrício Ávila Rodrigues

SOILS AND PLANT NUTRITION

159-168 Assessment of sugarcane harvesting residue effects on soil spectral behavior
José A. M. Demattê, Fabrício da Silva Terra, Rafael Otto, Raul Shiso Toma, Luiz Henrique Pereira, Alexandre Ferreira do Nascimento, Marco Antonio Melo Bortoletto

169-176 Sorption and desorption of indaziflam degradates in several agricultural soils
Diego Gonçalves Alonso, Rubem Silvério de Oliveira Jr., William Charles Koshiken, Kathleen Hall, Jamil Constantin, Suresh Mislankar

NOTE

177-183 Cytological aspects of incompatible and compatible interactions between rice, wheat and the blast pathogen Pyricularia oryzae
Leonardo Araújo, Juliana Moreira Soares, Marta Cristina Corsi de Filippi, Fabrício Ávila Rodrigues

184-188 Reaction of sources of resistance to white mold to microsatellite haplotypes of Sclerotinia sclerotiorum
Miller da Silva Lehner, Trazilbo José de Paula Júnior, Rogério Faria Vieira, Renan Cardoso Lima, Bruno de Almeida Soares, Raphael Alves Silva

REVIEW

189-197 Sulfites in beer: reviewing regulation, analysis and role
Luis F. Guido

199 Author Index

Instructions to Authors
INDEX OF AUTHORS

Alonso, D.G., 169
Alonzo, M.W., 103
Anjos, I.A., 115
Araujo, L., 150, 177
Avilez Ruiz, J.P., 103
Bendaou, N., 134
Bispo, W.M.S., 150
Bortoletto, M.A.M., 159
Constantin, J., 169
Delgado Pertínz, M., 103
Demattê, J.A.M., 159
Dinardo-Miranda, L.L., 115
Dolatowski, Z.J., 125
El-Haddoury, J., 134
Ferreira, C.F., 109
Filippi, M.C.C., 177
Fracasso, J.V., 115
Gesteira, A.S., 109
Girardi, E.A., 109
Guido, L.F., 189
Hall, K., 169
Henkrar, F., 134
Iraqi, D., 134
Izeppi, T.S., 115
Koskinen, W.C., 169
Lehner, M.S., 184
Lima, R.C., 184
Lopes, D.O.P., 115
Mislankar, S., 169
Moreira, W.R., 150
Nascimento, A.F., 159
Nsarellah, N., 134
Oliveira Jr., R.S., 169
Oliveira, M.C., 115
Otto, R., 159
Ouabbou, H., 134
Passos, O.S., 109
Paula Júnior, T.J., 184
Perecin, D., 115
Pereira, L.H., 159
Resende, M.D.V., 142
Riaz, S., 142
Rodrigues, F.A., 150, 177
Santos, M.G., 109
Silva, L.C., 150
Silva, R.A., 184
Soares Filho, W.S., 109
Soares, B.A., 184
Soares, J.M., 177
Soriano, J., 97
Terra, F.S., 159
Toma, R.S., 159
Udupa, S.M., 134
Viana, A.P., 142
Vieira Neto, J.G., 97
Vieira, R.F., 184
Walker, M.A., 142
Wojcjak, K.M., 125
Sorption and desorption of indaziflam degradates in several agricultural soils

Diego Gonçalves Alonso1, Rubem Silvério de Oliveira Jr.1*, William Charles Koskinen2, Kathleen Hall3, Jamil Constantin3, Suresh Mislankar4

1 State University of Maringá/Center for Advanced Studies in Weed Research (NAPD) – Agronomy Dept., Av. Colombo, 579 – 87020-900 – Maringá, PR – Brazil.
4 Bayer Crop Science LP, Environmental Fate and Exposure Assessment, RTP, 2 TW Alexander Drive – 27709 – Research Triangle Park, NC, USA.

*Corresponding author <rsojunior@uem.br>

Edited by: Silvia del Carmen Imhoff

Received March 13, 2015
Accepted July 20, 2015

ABSTRACT: Processes regulating pesticide fate in the environment are influenced by the physico-chemical properties of pesticides and soils. Sorption and desorption are important processes as they regulate the movement of pesticides in soil. Although sorption-desorption is widely studied for herbicides, studies involving their metabolites in soil are scarce. Sorption and desorption of indaziflam metabolites (indaziflam-triazinediamine (FDAT), indaziflam-triazine-indanone (ITI) and indaziflam-carboxilic acid (ICA)) were investigated in six Brazilian (BRA) soils and three United States (USA) soils with different physicochemical properties. The Freundlich equation described sorption of the metabolites for all soils ($R^2 > 0.98; 1/n ~ 1$). Sorption order ($K_f$) was ITI > ICA > FDAT. Mean values of $K_f$ oc were 453, 289, and 81 (BRA) and 444, 48, and 48 (USA) for metabolites ITI, ICA, and FDAT respectively. Desorption was hysteretic for all metabolites in all soils. These results suggest that these metabolites fall in the classification range of mobile to moderately mobile in soils.

Keywords: adsorption, hysteresis, metabolites, soil properties

Introduction

The fate and distribution of agrochemicals in the soil water environment are largely determined by sorption-desorption processes. Previous herbicide research has focused predominantly on the interaction of the parent compound with soil, while studies involving their metabolites are in short supply. Because data are unavailable for model input, it is often assumed that the metabolite sorption coefficient is equal to that of the parent compound. Studies on the metabolites of imidacloprid have demonstrated that this assumption can be inaccurate (Papiernik et al., 2006). For example, of three imidacloprid metabolites evaluated, two showed sorption higher than the parent compound and one lower (Cox et al., 1997; Papiernik et al., 2006). Alternatively, the idea that as a compound degrades, its byproducts are smaller and more polar, and, therefore, more mobile is also riddled with exceptions. The hydroxylated metabolites of atrazine (Clay and Koskinen, 1990) and florasulam (Krieger et al., 2000) show greater adsorption to soil than the parent compound. Experimental metabolite sorption data is, therefore, imperative for performing environmental assessments of herbicides. While information regarding herbicide metabolite behavior in soil is scarce overall, no data exist for the relatively new herbicide, indaziflam.

Indaziflam [N-[[1R,2S]-2,3-dihydro-2,6-dimethyl-1H-inden-1-yl]-6-[[1R]-1-fluoroethyl]-1,3,5-triazine-2,4-diamine] is a herbicide recently approved for use in the United States and is in the process of being released in Brazil. The main transformation chemicals from the environmental degradation of indaziflam are: indaziflam-triazine-indanone (ITI), indaziflam-carboxilic acid (ICA), indaziflam-hydroxyethyl, indaziflam-olefin, fluoroethyl-diaminotriazine (FDAT), and fluoroethyltriazinanedione. Indaziflam, ITI and ICA are cleaved at the N bond to form FDAT (from the triazine portion) and unidentified minor compounds (from the indazyl portion). Residues are ultimately converted into bound residues and CO$_2$ (Figure 1) [USEPA, 2010].

This study aimed to characterize sorption and desorption coefficients of three of the main metabolites of indaziflam (ITI, ICA and FDAT), for which there currently is no published information. Considering the diversity among the physicochemical properties of soils that can influence sorption-desorption, this study was carried out on a selection of tropical soils from Brazil (Oxisols) and glacial soils from the United States (Mollisols).

Materials and Methods

Soils

Nine various soils previously untreated with indaziflam were selected for this study: six Brazilian Oxisols and three United States Mollisols. Samples were collected from a depth of 0 to 10 cm depth, air-dried, and passed through a 2-mm sieve. Selected physicochemical properties of the soils appear in Table 1. The hydrometer method was used to determine sand and clay contents and soil pH was measured in a 1:2 soil/deionized water mixture. The soil organic carbon (OC) content was ascertained by loss on ignition (Nelson and Sommers, 1982).
Chemicals

Pure analytical standards of indaziflam-triazinediamine (FDAT) [6-[(1R)-1-fluoroethyl]-1,3,5-triazine-2,4-diamine], indaziflam-triazine-indanone (ITI) [N-[(1R,2S)-2,3-Dihydro-2,6-dimethyl-3-oxo-1H-inden-1-yl]-1,3,5-triazine-2,4-diamine] and indaziflam-carboxylic acid (ICA) [2S,3R]-3-[[4-amino-6-[(1R)-1-fluoroethyl]-1,3,5-triazin-2-yl]-amino]-2,3-dihydro-2-methyl-1H-indene-5-carboxylic acid, and respective radioactive analytical standards, 14C-labeled [indane-1,14C] ITI, and 14C-labeled [carboxyl-14C] ICA were graciously provided by Bayer Crop Science (Wuppertal, Germany). Solutions were prepared in CaCl2 (0.01 N) to concentrations of 0.06, 0.19, 0.64, and 1.91 µmol L−1 (FDAT), 0.03, 0.09, 0.3, and 0.95 µmol L−1 (ITI), and 0.03, 0.09, 0.31, and 0.92 µmol L−1 (ICA) with nonradioactive analytical standards. Radioactive analytical standards were added to provide ~83 Bq mL−1 of radioactivity to solutions. Final solutions contained less than 0.4 % methanol and were stored in foil-covered flasks in the dark at 4 °C.

Sorption

Sorption studies were performed in triplicate using the batch equilibration method. Preliminary studies (Alonso et al., 2011) showed that to sorb 20-80 % of the initial chemical in solution on the soil (for greater analytical accuracy), a ratio of 4 g soil to 10 mL (4:10) solution was acceptable for the ITI metabolite, whereas for FDAT and ICA, the soil:solution ratio was 10 g to 10 mL (1:1) for each sample. Glass centrifuge tubes (35-mL) with Teflon lined caps containing soil and radioactive solutions were mixed using a vortex mixer (30 s) and shaken for 24 h on a tabletop shaker (20 ± 2 °C) in the dark. Preliminary kinetic studies (Alonso et al., 2011) showed that equilibrium was reached in < 24 hrs. Sample tubes were then centrifuged (20 min at 370 g) and 3-mL of supernatant was removed from each sample for analysis. A 1-mL aliquot of the removed supernatant was mixed with 5 mL of a scintillation cocktail (EcoLite, cocktail, ICN Biomedicals, Costa Mesa, CA) and the concentration of 14C of each metabolite in solution was determined by liquid scintillation counting with a Packard 1500 counter (Packard Instruments, Downer Grove, IL). The amount of chemical sorbed to the soil was calculated as the difference between the initial solution concentration and the supernatant concentration.
after equilibration. In preliminary studies, degradation of the chemical during equilibration was not observed, neither was sorption in blank tubes containing solution without soil. Data were fit to the Freundlich equation to obtain sorption coefficients.

**Desorption**

Immediately following the sorption experiments, desorption experiments were performed for each metabolite at the lowest and highest initial concentrations: 0.06 and 1.91 µmol L\(^{-1}\) for FDAT, 0.03 and 0.95 µmol L\(^{-1}\) for ITI, and 0.03 and 0.92 µmol L\(^{-1}\) for ICA on all nine soils. Desorption was determined by replacing the 3-mL aliquot of removed supernatant for sorption analysis with 3 mL of 0.01 N CaCl\(_2\). The soils were then re-suspended using a vortex mixer, shaken for 24 h, and centrifuged. The supernatants were analyzed as previously described and again replaced with CaCl\(_2\). This was repeated for a total of three desorption steps. All studies were done in triplicate.

**Sorption and Desorption Model**

Sorption coefficients \(K_f\) and \(1/n\) were calculated from the slope and intercept of the log form of the Freundlich equation: Log \([Cs]\) = log \(K_f\) + \(1/n\) log \([Ce]\); where \([Cs]\) is the concentration [µmol kg\(^{-1}\)] of the metabolite sorbed onto soil after equilibration and \([Ce]\) is the metabolite concentration [µmol L\(^{-1}\)] after equilibration. Sorption coefficients were normalized to soil OC contents to give \(K_{foc}\) [L kg\(^{-1}\)], where \(K_{foc} = [K_f] / \% OC)*100. The hysteresis coefficient, \(H\), for the sorption-desorption isotherms was determined using the formula \(H = (1/1/n_{desorption})(1/1/n_{sorption})\), where \(1/n_{sorption}\) and \(1/n_{desorption}\) are the Freundlich slopes from the sorption and desorption isotherms, respectively (Barriuso et al., 1994).

Sorption distribution coefficients \(K_d\) [L kg\(^{-1}\)], where \(K_d = [Cs] / [Ce]\), were also calculated for the lowest and highest initial solution concentrations and normalized to soil OC contents to give \(K_{doc}\) [L kg\(^{-1}\)], where \(K_{doc} = [K_d] / \% OC)*100. \(K_d\) and \(K_{doc}\) values were used to compare sorption between soils at initial solution concentrations with sorption calculated after the third desorption step. The concentration of metabolite remaining in the 0.01 N CaCl\(_2\) solution after the third desorption step was taken as \([C_e]\), and the concentration of metabolite sorbed, \([C_s]\), was assumed to equal the initial concentration minus the amount remaining in the solution and the amount removed in the previous desorption steps.

**Statistical Analysis**

For each metabolite, Pearson Product Moment Correlation analyses were performed charting the sorption coefficients against selected soil physicochemical properties using the SigmaPlot (SysStat’ for Windows, Version 10). For all correlation analyses, soils were divided into two groups, tropical soils (BRA) and glacial soils (USA). Linear regression analyses were performed using the same software package.

**Results and Discussion**

**Sorption**

The empirical Freundlich equation described the sorption behavior of the metabolites FDAT (Table 2, Figure 2), ITI (Table 3, Figure 3), and ICA (Table 4, Figure 4) for all soils \((R^2 > 0.98)\). Average \(1/n\) values were 0.99 for FDAT, and 0.95 for ITI and ICA indicating sorption was minimally dependent on initial solution concentration. As \(1/n\) values did not differ significantly from one another, \(K_f\) values were used for comparisons. In general, the order of sorption magnitude for the indaziflam metabolites in the present study was FDAT < ICA < ITI. \(K_f\) values for FDAT, the least sorbed metabolite, were 0.14 to 3.0 in Oxisols and 0.63 to 0.85 in Mollisols. ICA \(K_f\) values of Oxisols ranged from 0.51 to 13.1, whereas...
responding soil (Alonso et al., 2011), which supports the general observation that degradates are more polar and soluble than parent chemicals, and subsequently less sorbed.

Table 2 – Freundlich sorption parameters for indaziflam-triazinediamine (FDAT) metabolite of indaziflam for soils studied.

<table>
<thead>
<tr>
<th>Soil</th>
<th>$K_f$</th>
<th>$K_{f,oc}$</th>
<th>$1/n$ sorption</th>
<th>$R^2$</th>
<th>Init. Conc. Desorption</th>
<th>$1/n$ desorption</th>
<th>$H$</th>
<th>$K_{f,oc}/K_f$</th>
</tr>
</thead>
<tbody>
<tr>
<td>BRA 1</td>
<td>0.93 (0.93-0.94)</td>
<td>58 (58-59)</td>
<td>0.99 ± 0.0</td>
<td>1.0</td>
<td>1.91</td>
<td>0.41</td>
<td>0.42</td>
<td>1.54</td>
</tr>
<tr>
<td>BRA 3</td>
<td>0.14 (0.13-0.15)</td>
<td>28 (26-30)</td>
<td>1.0 ± 0.04</td>
<td>0.9963</td>
<td>1.91</td>
<td>0.12</td>
<td>0.12</td>
<td>2.38</td>
</tr>
<tr>
<td>BRA 7</td>
<td>1.18 (1.16-1.19)</td>
<td>57 (57-58)</td>
<td>0.97 ± 0.01</td>
<td>0.9999</td>
<td>1.91</td>
<td>0.26</td>
<td>0.26</td>
<td>1.66</td>
</tr>
<tr>
<td>BRA 8</td>
<td>0.5 (0.47-0.54)</td>
<td>23 (22-25)</td>
<td>1.0 ± 0.04</td>
<td>0.9975</td>
<td>1.91</td>
<td>0.44</td>
<td>0.44</td>
<td>1.62</td>
</tr>
<tr>
<td>BRA 11</td>
<td>3.0 (2.64-3.41)</td>
<td>283 (249-322)</td>
<td>1.0 ± 0.05</td>
<td>0.9957</td>
<td>1.91</td>
<td>0.67</td>
<td>0.67</td>
<td>1.05</td>
</tr>
<tr>
<td>BRA 12</td>
<td>0.21 (0.21-0.22)</td>
<td>35 (34-36)</td>
<td>1.02 ± 0.02</td>
<td>0.9996</td>
<td>1.91</td>
<td>0.29</td>
<td>0.29</td>
<td>2.05</td>
</tr>
<tr>
<td>USA 1</td>
<td>0.63 (0.61-0.65)</td>
<td>29 (28-30)</td>
<td>0.98 ± 0.01</td>
<td>0.9997</td>
<td>1.91</td>
<td>0.44</td>
<td>0.44</td>
<td>1.54</td>
</tr>
<tr>
<td>USA 2</td>
<td>0.90 (0.88-0.92)</td>
<td>82 (80-83)</td>
<td>0.99 ± 0.01</td>
<td>0.9998</td>
<td>1.91</td>
<td>0.48</td>
<td>0.48</td>
<td>1.51</td>
</tr>
<tr>
<td>USA 3</td>
<td>0.85 (0.84-0.86)</td>
<td>34 (33-34)</td>
<td>0.95 ± 0.01</td>
<td>0.9999</td>
<td>1.91</td>
<td>0.44</td>
<td>0.44</td>
<td>1.48</td>
</tr>
</tbody>
</table>

Table 3 – Freundlich sorption parameters for indaziflam-triazine-indanone (ITI) metabolite of indaziflam for soils studied.

<table>
<thead>
<tr>
<th>Soil</th>
<th>$K_f$</th>
<th>$K_{f,oc}$</th>
<th>$1/n$ sorption</th>
<th>$R^2$</th>
<th>Init. Conc. Desorption</th>
<th>$1/n$ desorption</th>
<th>$H$</th>
<th>$K_{f,oc}/K_f$</th>
</tr>
</thead>
<tbody>
<tr>
<td>BRA 1</td>
<td>10.0 (7.34-13.61)</td>
<td>625 (459-851)</td>
<td>0.95 ± 0.08</td>
<td>0.9853</td>
<td>0.95</td>
<td>0.05</td>
<td>0.06</td>
<td>1.97</td>
</tr>
<tr>
<td>BRA 3</td>
<td>1.96 (1.79-2.14)</td>
<td>392 (358-428)</td>
<td>0.92 ± 0.03</td>
<td>0.9976</td>
<td>0.95</td>
<td>0.31</td>
<td>0.33</td>
<td>1.66</td>
</tr>
<tr>
<td>BRA 7</td>
<td>14.46 (13.22-15.83)</td>
<td>706 (645-772)</td>
<td>1.02 ± 0.02</td>
<td>0.9989</td>
<td>0.95</td>
<td>0.18</td>
<td>0.18</td>
<td>1.42</td>
</tr>
<tr>
<td>BRA 8</td>
<td>4.26 (4.02-4.52)</td>
<td>196 (185-208)</td>
<td>0.98 ± 0.02</td>
<td>0.9992</td>
<td>0.95</td>
<td>0.26</td>
<td>0.26</td>
<td>1.59</td>
</tr>
<tr>
<td>BRA 11</td>
<td>4.24 (4.13-4.35)</td>
<td>400 (390-410)</td>
<td>0.94 ± 0.01</td>
<td>0.9998</td>
<td>0.95</td>
<td>0.50</td>
<td>0.53</td>
<td>1.24</td>
</tr>
<tr>
<td>BRA 12</td>
<td>2.46 (2.32-2.60)</td>
<td>403 (381-425)</td>
<td>0.92 ± 0.02</td>
<td>0.9991</td>
<td>0.95</td>
<td>0.24</td>
<td>0.26</td>
<td>0.70</td>
</tr>
<tr>
<td>USA 1</td>
<td>9.19 (8.25-10.24)</td>
<td>422 (378-470)</td>
<td>0.94 ± 0.03</td>
<td>0.9990</td>
<td>0.95</td>
<td>0.07</td>
<td>0.08</td>
<td>1.85</td>
</tr>
<tr>
<td>USA 2</td>
<td>4.72 (4.48-4.96)</td>
<td>429 (408-451)</td>
<td>0.93 ± 0.02</td>
<td>0.9994</td>
<td>0.95</td>
<td>0.22</td>
<td>0.24</td>
<td>1.62</td>
</tr>
<tr>
<td>USA 3</td>
<td>12.14 (12.60-12.71)</td>
<td>482 (460-504)</td>
<td>0.91 ± 0.01</td>
<td>0.9997</td>
<td>0.95</td>
<td>0.16</td>
<td>0.18</td>
<td>1.45</td>
</tr>
</tbody>
</table>

$K_f$ values for Mollisols ranged from 0.43 to 1.56. ITI $K_f$ values in Oxisols ranged from 1.96 to 14.5 and from 4.72 to 12.1 in Mollisols. All metabolites showed much lower sorption than the parent compound in the corresponding soil (Alonso et al., 2011), which supports the general observation that degradates are more polar and soluble than parent chemicals, and subsequently less sorbed.
Table 4 – Freundlich sorption parameters for indaziflam-carboxylic acid (ICA) metabolite of indaziflam for soils studied.

<table>
<thead>
<tr>
<th>Soil</th>
<th>$K_f$</th>
<th>$K_{oc}$</th>
<th>$1/n$ sorption</th>
<th>$R^2$</th>
<th>Init. Conc. desorption</th>
<th>$1/n$ desorption</th>
<th>$H$</th>
<th>$K_{oc3}/K_{oc}$</th>
</tr>
</thead>
<tbody>
<tr>
<td>BRA 1</td>
<td>1.78 (1.44-2.20)</td>
<td>111 (90-137)</td>
<td>0.83 ± 0.08</td>
<td>0.9831</td>
<td>0.92</td>
<td>0.13</td>
<td>0.15</td>
<td>1.89</td>
</tr>
<tr>
<td>BRA 3</td>
<td>0.51 (0.49-0.53)</td>
<td>102 (97-107)</td>
<td>0.96 ± 0.02</td>
<td>0.9993</td>
<td>0.92</td>
<td>0.40</td>
<td>0.41</td>
<td>1.63</td>
</tr>
<tr>
<td>BRA 7</td>
<td>13.08 (12.89-13.27)</td>
<td>638 (629-648)</td>
<td>0.99 ± 0.00</td>
<td>1.0</td>
<td>0.92</td>
<td>0.09</td>
<td>0.09</td>
<td>1.44</td>
</tr>
<tr>
<td>BRA 8</td>
<td>6.51 (6.18-6.85)</td>
<td>300 (285-316)</td>
<td>1.02 ± 0.02</td>
<td>0.9995</td>
<td>0.92</td>
<td>0.27</td>
<td>0.26</td>
<td>1.03</td>
</tr>
<tr>
<td>BRA 11</td>
<td>5.10 (5.04-5.16)</td>
<td>481 (476-487)</td>
<td>1.01 ± 0.00</td>
<td>1.0</td>
<td>0.92</td>
<td>0.21</td>
<td>0.21</td>
<td>1.14</td>
</tr>
<tr>
<td>BRA 12</td>
<td>0.62 (0.57-0.67)</td>
<td>101 (93-110)</td>
<td>0.93 ± 0.04</td>
<td>0.9962</td>
<td>0.92</td>
<td>0.37</td>
<td>0.40</td>
<td>1.65</td>
</tr>
<tr>
<td>USA 1</td>
<td>0.94 (0.92-0.96)</td>
<td>43 (42-44)</td>
<td>0.93 ± 0.01</td>
<td>0.9998</td>
<td>0.92</td>
<td>0.13</td>
<td>0.13</td>
<td>1.99</td>
</tr>
<tr>
<td>USA 2</td>
<td>0.43 (0.41-0.46)</td>
<td>39 (38-41)</td>
<td>0.93 ± 0.02</td>
<td>0.9992</td>
<td>0.92</td>
<td>0.21</td>
<td>0.23</td>
<td>2.00</td>
</tr>
<tr>
<td>USA 3</td>
<td>1.56 (1.47-1.66)</td>
<td>62 (58-66)</td>
<td>0.98 ± 0.02</td>
<td>0.9991</td>
<td>0.92</td>
<td>0.31</td>
<td>0.31</td>
<td>1.55</td>
</tr>
</tbody>
</table>

1Numbers in parentheses are confidence intervals ($K_f$, $K_{oc}$) or standard deviations of the means ($1/n$).

Figure 4 – Freundlich isotherms fit to describe sorption (solid symbols) and desorption (open symbols) of indaziflam-carboxylic acid (ICA) at concentrations of 0.03 and 0.92 µmol L$^{-1}$ for soils BRA3, BRA7, USA2, and USA3.

Sorption of FDAT was lower as compared to indaziflam, presumably due to differences in hydrophobic interactions. Using a theoretical pKa for FDAT of 4.10 (Chemaxon, 2014), a higher %age of cationic molecules would be expected as compared to indaziflam (pKa = 3.5), and greater sorption. However, the pKa is not the only factor that affects sorption. There was no correlation ($p < 0.05$) between low FDAT $K_f$ values and the physicochemical properties (ie. pH and %age clay) of the soils. Although there was no significant correlation between $K_f$ and OC, the normalization of $K_f$ for organic carbon (% OC) reduced the variability of sorption coefficients. Variation range in $K_{oc3}$ was ~12x in contrast to the ~21x variation for $K_{oc}$.

The ICA $K_f$ values of Oxisols had significant positive correlation ($p < 0.05$) with % OC ($r = 0.76**$), and an equation describing $K_f$ as a function of % OC was calculated where $K_f = -2.1738 + 5.0867 \% OC$ ($R^2 = 0.99$). ICA sorption was not correlated with clay content or pH of these soils. In the Mollisols, significant negative correlations ($p < 0.05$) were observed for ICA $K_f$ values with pH ($r = -0.93**$) and clay content ($r = 0.99**$). Linear equations describing $K_f$ as a function of soil pH ($K_f = 4.0454 - 0.411 \text{pH}$, $R^2 = 0.87$), clay contents ($K_f = 3.6604 - 0.1413 \% \text{Clay}$, $R^2 = 0.99$), and % OC ($K_f = -0.4103 + 0.7174 \% \text{OC}$, $R^2 = 0.88$) were calculated. Although not significant, the normalization of $K_f$ or $K_{oc}$ for organic carbon content reduced sorption variability between soils by approximately half, from ~30 to ~16. The low ICA sorption is presumably related to the anionic character of the metabolite at the pH of the suspensions. The anionic acid group is repelled by the negative charge of Oxisols, or weakly retained as the molecular species by most soil and sediment components in glacial soils (FAO, 2000). According to calculations, pKa varied from 3.70 to 4.36, depending on the group, triazine ring or the acid group (Chemaxon, 2014).

$K_{oc}$ was positively correlated ($p < 0.05$) with organic carbon (% OC) ($r = 0.68**$) and with clay content (% Clay) ($r = 0.74**$) of the Oxisols. Linear equations describing $K_f$ as a function of carbon content ($K_f$,
= 0.0171 + 4.6889 % OC, $R^2 = 0.46$) and $K_f$ as a function of clay content ($K_f = 1.4215 + 0.1342$ % Clay, $R^2 = 0.54$) were calculated. The poor fit observed for both cases is directly related to low sorption by soil BRA8. Although its physicochemical characteristics are similar to soil BRA7, its $K_f$ was ~3 times lower. Similar behavior was observed for the sorption of the parent compound [indaziflam] for the same soil (Alonso et al., 2011). It is surprising that there was no correlation between sorption and pH. The theoretical pKa for triazine indanone is 4.10 (Chemaxon, 2014), and the %age cationic molecules at a given pH would be expected to contribute to sorption. However, as was mentioned above, the pKa is not the only factor that affects the sorption.

For the three Mollisols studied, there were significant incidences of correlation between the ITI Freundlich sorption coefficients and soil pH, % OC and % clay. The pH ($r = -0.85**$) and clay content ($r = -0.99**$) were negatively correlated with $K_f$, whereas the organic carbon content correlated positively with the $K_f$ for these soils ($r = 0.98**$). Linear equations describing $K_f$ as a function of soil pH ($K_f = 27.2005 - 2.48$ pH, $R^2 = 0.72$), clay contents ($K_f = 26.3058 - 0.9275$ % Clay, $R^2 = 0.99$), and % OC ($K_f = -0.9101 + 4.9621$ % OC, $R^2 = 0.97$) were significant. The negative correlation observed between $K_f$ and clay content is possibly a result of the positive correlation between $K_f$ and the organic carbon content of these soils overshadowing the contribution of the clay. The normalization of both $K_f$ and $K_{f,oc}$ for % OC [$K_{f,oc}$] reduced the coefficients’ range of variability from ~7.5x to ~4x.

When the sorption of metabolites is very low, such as is the case with FDAT, the chemical properties of the soil will bear little or no relation to the mobility of the compound. On the other hand, when the metabolite presents higher sorption, the main chemical property contributing to increased sorption is the carbon content, which showed significant correlation with the sorption of these compounds in most cases. This correlation is reiterated by the fact that the normalization of $K_f$ and $K_{f,oc}$ to carbon contents $K_{f,oc}$ and $K_{f,oc}$ respectively, reduced the variability between the coefficients.

The main soil variable that was negatively correlated with the sorption of metabolites was pH. The inverse relationship between pH and sorption suggests that the compounds may be partially ionized in the soil solution. ITI and FDAT have N-groups, which may be protonated at low pH, while ICA may be dissociated at high pH. On the other hand, compounds that are not ionized in the soil tend not to show any correlation between sorption and pH.

The inconsistent effects of clay content on the sorption of metabolites, showing both positive [Oxisols] and negative correlation [Mollisols], suggests that the influence of the organic fraction and pH of these soils are the main characteristics affecting sorption. In Mollisols organic carbon was inversely related to clay contents and pH. Therefore, sorption increased with the increase in % OC and the decrease in pH, which resulted in apparent negative correlation between $K_f$ and the clay fraction of these soils. Negative correlation between $K_f$ and the clay fraction of these soils was also observed for the parent compound [indaziflam] (Alonso et al., 2011).

Desorption

Desorption of the three metabolites was found to be hysteretic for all soils studied, $1/n_{desorption} < 1/n_{sorption}$ (Tables 2, 3 and 4). Figures 2, 3 and 4 show representative desorption isotherms of the metabolites in four soils at the lowest and the highest initial concentrations used in the study. Hysteresis coefficients, $H$, ranged from 0.27 to 0.49 (in 0.06 $\mu$mol L$^{-1}$) and from 0.12 to 0.67 (in 1.91 $\mu$mol L$^{-1}$) for Oxisols, and from 0.46 to 0.56 (in 0.06 $\mu$mol L$^{-1}$) and from 0.45 to 0.49 (in 1.91 $\mu$mol L$^{-1}$) for Mollisols (Table 2).

It is difficult to ascribe the degree of hysteresis to a particular soil property. Although $H$ showed a significant positive correlation ($p < 0.05$) with Oxisol $K_f$ values ($r = 0.71**$) [described by the equation $H = 0.2784 - 0.0998 K_f$, $R^2 = 0.50$], no correlation was found between $H$ and the physicochemical characteristics of the Oxisols in this study. In contrast, in the Mollisols negative correlation was observed between $H$ and % OC ($r = -0.78**$) and positive correlation between $H$ and % Clay ($r = 0.66**$) and between $H$ and $K_f$ ($r = 0.61**$). Multiple correlation was described by the following linear equations: $H = 0.5739 - 0.0477$ % OC, $R^2 = 0.60$; $H = 0.3392 + 0.0075$ % Clay, $R^2 = 0.43$; and $H = 0.3272 + 0.1947 K_f$, $R^2 = 0.38$. No correlation was found between $H$ and pH of the Mollisols.

ITI hysteresis coefficients [$H$] for Oxisols ranged from 0.02 to 0.46 (in 0.03 $\mu$mol L$^{-1}$) and from 0.06 to 0.53 (in 0.95 $\mu$mol L$^{-1}$). For Mollisols, $H$ ranged from 0.03 to 0.13 (in 0.03 $\mu$mol L$^{-1}$) and from 0.08 to 0.24 (in 0.95 $\mu$mol L$^{-1}$) [Table 2]. There was significant negative correlation ($p < 0.05$) between $H$ and Oxisol clay contents [$r = -0.67**$], described by the equation $H = 0.3782 - 0.0039$ % Clay, $R^2 = 0.45$. No correlation was found between $H$ and $K_f$, or between $H$ and the other reported characteristics of the soils.

ICA hysteresis coefficients, $H$, ranged from 0.10 to 0.72 (in 0.03 $\mu$mol L$^{-1}$) and from 0.09 to 0.41 (in 0.92 $\mu$mol L$^{-1}$) for Oxisols and from 0.14 to 0.25 (in 0.03 $\mu$mol L$^{-1}$) and 0.13 to 0.31 (in 0.92 $\mu$mol L$^{-1}$) for Mollisols. There was significant negative correlation ($p < 0.05$) between $H$ and % OC [$r = -0.74**$], % Clay [$r = -0.76**$], and $K_f$ [$r = -0.63**$] for Oxisols, described by the following equations: $H = 0.5537 - 0.1930$ % OC, $R^2 = 0.54$; $H = 0.4865 - 0.0053$ % Clay, $R^2 = 0.58$; and $H = 0.4102 - 0.0247 K_f$, $R^2 = 0.40$. No correlation was found between $H$ and Oxisol pH$_5$, nor was correlation found between $H$ and the physicochemical characteristics or $K_f$ of Mollisols.

Potential Mobility

As a result of the lower sorption of the FDAT, ITI, and ICA metabolites compared to the parent compound,
it is expected that their mobility will be higher than indaziflam. Based on the criteria established by the Food and Agriculture Organization of the United Nations (FAO) [FAO, 2000], FDAT $K_{f-oc}$ variation found in this study [Table 2], from 23 to 283 L kg$^{-1}$ for tropical Oxisols and from 29 to 82 L kg$^{-1}$ for glacial Mollisols, would be classified as FDAT as mobile in all soils studied except BRA11, in which it is considered moderately mobile. Based on $K_{f-oc}$ values [Table 3], ITI would be classified as moderately mobile in Oxisols [$K_{f-oc} = 196-706$ L kg$^{-1}$] and Mollisols [$K_{f-oc} = 422-482$ L kg$^{-1}$]. ICA $K_{f-oc}$ values for Oxisols ranged from 101 to 638 L kg$^{-1}$ and from 39 to 62 L kg$^{-1}$ for Mollisols [Table 3]. Thus, the metabolite ICA would be classified as mobile to moderately mobile in Oxisols, and considered mobile in Mollisols [FAO, 2000].

Although sorption initially regulates soil-pesticide interactions, desorption also influences the potential mobility of these molecules and the risk of environmental contamination. Desorption represents the magnitude of the sorbed fraction that can return to the solution by reaching a new equilibrium. This typically occurs when a portion of the chemical initially present in solution is shifted in the soil profile or removed by adsorption, degradation, leaching, or volatilization. Therefore, to fully understand compound availability as a function of time, it is necessary to account for the desorbed fraction re-entering the solution.

Desorption of the three metabolites is hysteretic, which means it does not occur to the same degree as sorption, $1/n_{sorption} > 1/n_{desorption}$, and a large portion of sorbed metabolites is not easily desorbed or is desorbed slowly. Desorption hysteresis is caused by a variety of mechanisms for a large number of herbicide-soil systems [Wauchope et al., 2002]. Weber et al. [1998] suggest that hysteresis can most likely be attributed to slow desorption and retention of sorbed molecules within highly condensed organic matrices. The highest coefficients of hysteresis were found for FDAT, indicating that along with being the least sorbed, this metabolite also has the highest desorption potential among those studied as a result of being partially protonated in the soil solution. The FAO mobility classification system [FAO, 2000] does not take this desorption hysteresis into account. Because the hysteretic behavior of chemicals reduces their mobility, the potential of contamination is likely overestimated.

A variety of transport models have been used to predict the potential leaching depth of a chemical in soil. The sorption coefficient, $K_d$, is the most commonly used retardation factor and the depth of leaching is directly proportional to the magnitude of $K_d$. This coefficient does not take into account hysteresis, if it occurs during desorption. For FDAT, use of the sorption $K_d$ would overpredict the depth of leaching by a factor of 1.5 to 2 as compared to using the sorption coefficients for the third desorption equilibration ($K_{des3}$) as indicated by the $K_{pread}/K_{d}$ ratio [Table 2]. Similarly, depending on the soil, potential depth of leaching could be overestimated by a factor of 2 for both ITI and ICA.

The evaluation of the potential leaching of these metabolites to ground water may vary according to environmental conditions, particularly with regard to their persistence in the soil. The more persistent these chemicals are, the greater the likelihood of them becoming major contaminants. Thus, complementary studies to determine the half-life of these compounds in the soil should be carried out with the aim of fully understanding their potential to become contaminants.

In summary, the three metabolites showed distinct behavior in terms of sorption in the soil, which suggests that they should be treated as isolated molecules. The three compounds are less sorbed than the parent compound, which indicates that they have a higher leaching potential in the soil, especially in those with low carbon content and high pH. Hysteresis was observed in desorption experiments indicating an imbalance in the sorption-desorption process, which may signify the presence of strongly sorbed fractions and the formation of bound residues.

**Acknowledgments**

We thank CNPq and CAPES (Brazil) for financial support for this research. We also thank Bayer for supplying the analytical and radioactive materials; Luis Ávila (UFPel, RS), Hugo Dan (FESURV, GO), and Miriam Inoue [UNEMAT, MT] for their help in collecting soil samples.

**References**


